



Cortisone
CLIA kit
(96 Tests)

Zellbio GmbH (Germany)

CAT No. ZX-66104-96

www.zellx.de

Sample Types Validated for:

Serum, Plasma, Urine, Saliva, Dried Fecal Extracts, Tissue Culture Media

!!! Caution: This product is for Research Use Only. Not for *in vitro* Diagnostics !!!

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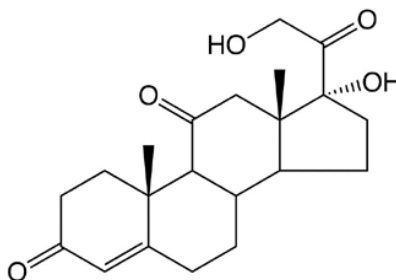
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Please read this insert completely prior to using the product.

Introduction

Background

Cortisone (C₂₁H₂₈O₅, compound E) was identified and extracted from bovine suprarenal gland tissue by Mason, Myers and Kendall in 1936. The presence of multiple cortin-like compounds led the authors to speculate that the study of Compound E, which had the qualitative but not quantitative activity of cortin, would reveal the nature of cortin. Compound E and the more active glucocorticoid, compound F, are now called Cortisone and Cortisol, respectively. The concentrations of these two glucocorticoids vary due to the activity of two enzymes 11 β -hydroxysteroid dehydrogenases 1 and 2 (11 β -HSD). While most tissues have the ability to express either enzyme, 11 β -HSD1 is found primarily in the liver where it converts Cortisone to Cortisol, whereas 11 β -HSD2 is found in tissues such as the kidney where Cortisol receptor binding is required. 11 β -HSD2 deactivates Cortisol to Cortisone, and prohibits receptor activation. This glucocorticoid “shuttle” helps to initiate and regulate the anti-inflammatory response, making Cortisone one of the modern “wonder drugs”. Monitoring the ratio of Cortisone/Cortisol has applications in diabetes, obesity, metabolic syndrome, osteoporosis, and chronic fatigue syndrome in addition to adrenal diseases. Cortisone and Cortisol concentrations exhibit a predictable diurnal pattern and can be measured in extracted dried feces, or in serum, plasma, saliva and urine.



Assay principle

The ZELLX® Cortisone Immunoassay kit is designed to quantitatively measure Cortisone present in serum, plasma, urine, saliva, extracted dried fecal samples, and tissue culture media samples. A Cortisone stock solution is provided to generate a standard curve for the assay and all samples should be read off the standard curve.

The kit includes a 96-well plate that is pre-coated with a secondary anti-rabbit antibody. The function of this antibody is to capture the rabbit anti-Cortisone antibody bound to Cortisone conjugate (peroxidase-labeled) and hold this complex to the plate during the subsequent detection steps. The Cortisone-conjugate (labeled) and the sample Cortisone (unlabeled) compete for binding to the mouse antibody. After 2 hours of incubation, the chemiluminescent substrate is added to react with the peroxidase-labeled antibody-antigen conjugate to produce light. The generated light can be measured in a microtiter plate reader capable of reading luminescence. The lower the amount of Cortisone in the sample, the stronger the signal due to more labeled cortisone bound to the well.

General information

Materials supplied in the Kit

Component	Quantity
Cortisone Standard (1000 ng/mL)	25 µL
Cortisone Antibody	2.6 mL
Cortisone Conjugate Concentrated	700 µL
Conjugate Diluent	2.6 mL
Assay Buffer Concentrate (5x)	11 mL
Wash Buffer Concentrate (20x)	25 mL
Dissociation Reagent	1 mL
Substrate A	5.6 mL
Substrate B	5.6 mL
Coated Clear 96-Well Plate & Sealer	1 plate

Storage instruction

All reagents should be stored at 4° C until the expiration date of the kit.

Materials required but not supplied

Deionized water (diH₂O)

Phosphate Buffer Saline (PBS)

Microplate reader capable of reading glow chemiluminescence.

Note: All luminometers read Relative Light Units (RLU). These RLU readings will vary with brand or model of plate reader. The number of RLUs obtained depends on the sensitivity and gain of the reader used. If you are not sure how to properly configure your reader, contact your plate reader manufacturer or carry out the following protocol:

Dilute 5 µL of the Conjugate Working Solution into 995 µL of deionized water. Pipet 5 µL of diluted conjugate into a white well and add 100 µL of prepared CLIA substrate. This well will give an intensity slightly above the maximum binding for the assay. Adjust the gain or sensitivity so that your reader is giving close to the maximum signal.

Microplate shaker, Centrifuge, Vortex mixer

Precision pipettes, multichannel/repeater pipettes and disposable pipette tips

For Dried Fecal Sample:

ACS Grade Ethanol

Glass test tubes

Precautions

This kit should only be used by qualified personnel who have had laboratory safety instruction. The complete insert should be read and understood before attempting to use the product.

General remarks

- Equilibrate all kit components to room temperature (RT) 30 minutes before use.
- The instruction must be strictly followed.
- The reading of Microplate/ELISA reader must be set at the appropriate wavelength.
- Pipette tips should not be used more than once in order to avoid cross contamination.
- Reagents of different batches should not be mixed or used after their expiration dates.
- The antibody-coated plate needs to be stored desiccated. The silica gel pack included in the foil ziploc bag will keep the plate dry. The silica gel pack will turn from blue to pink if the ziploc has not been closed properly.
- This kit utilizes a peroxidase-based readout system. Buffers, including Wash Buffers from other manufacturers, containing sodium azide will inhibit color production by the enzyme. Make sure all buffers used for samples are azide-free. Ensure that any plate washing system is rinsed well with deionized water prior to using the supplied Wash Buffer.

Assay protocol

Reagent preparation

- i. **Assay Buffer:** Prepare a 1:5 dilution of Assay Buffer Concentrate with diH₂O (1 part Assay Buffer Conc. with 4 parts diH₂O), and mix well. Assay Buffer can be stored at 4°C for up to 3 months.
- ii. **Wash Buffer:** Prepare a 1:20 dilution of Wash Buffer Concentrate with diH₂O (1 part Wash Buffer Conc. with 19 parts diH₂O), and mix well. Assay Buffer can be stored at room temperature for up to 3 months.

- iii. **Conjugate Working Solution:** Prepare a 1:4 dilution of Cortisone Conjugate Concentrated with Conjugate Diluent (1 Part of Cortisone Conjugate Concentrated with 3 part of Conjugate Diluent), Conjugate Working Solution can be stored at 4°C for up to 1 month.
- iv. **Chemiluminescent Substrate:** Mix one part of the Substrate A with one part of Substrate B in a brown bottle. Chemiluminescent Substrate can be stored at 4°C for up to 1 month.

Sample preparation

Samples containing visible particulate should be centrifuged prior to conducting the assay. Moderate to severely hemolyzed samples should not be used for this assay.

Since Cortisone is identical across all species, it is expected that this kit can measure Cortisone in human and other species.

Dissociation Reagent must be used only with Serum and Plasma samples.

All samples and standards must be used within 2 hours of preparation or must be stored at ≤ -20 for later analysis.

I. Serum, Plasma:

- Collect the fresh serum or plasma with heparin or EDTA.
- Centrifuge at 600 g for 10 min at 4°C.
- Separate the serum or plasma from the red blood cells, and transfer into fresh tubes.
- Serum and plasma samples need to be treated with the supplied Dissociation Reagent. Adding this reagent will yield the total Cortisone concentration in serum or plasma.
- Allow the Dissociation Reagent to warm completely to Room Temperature before use.
- Add 5 μ L of Dissociation Reagent into 1 mL Eppendorf tubes.
- Add 5 μ L of serum or plasma to the Dissociation Reagent in the tube, vortex gently and incubate at room temperature for at least 5 minutes.
- Add 490 μ L of Assay Buffer to the tube.
- This 1:100 dilution can be diluted further with Assay Buffer. Final serum and plasma dilutions should be $\geq 1:100$.

II. Dried Fecal Sample:

- Ensure that the sample is completely dry, and powder the sample to improve extraction recovery. Remove any large particles if possible.
- Weigh out ≥ 0.2 gram of dried fecal solid into a tube. Samples can be dried by passive drying, gentle heating ($\leq 60^{\circ}\text{C}$), or freeze-drying (lyophilization).
- Add 1 mL of ethanol per 0.1 gram of solid fecal sample (100 mg/mL) and seal.
- Shake strongly for at least 30 minutes.
- Centrifuge at 5000 rpm at 4°C for 15 minutes and collect supernatant in a clean tube. This material can be stored at $\leq -20^{\circ}\text{C}$ for at least a month if properly sealed.

- **Note:** Samples containing low levels of analyte can be concentrated by drying down the extract and resuspension in a reduced volume of Assay Buffer.
- Supernatant should be diluted $\geq 1:5$ by taking one part of sample and adding 4 or more parts of Assay Buffer.
- Vortex well and allow to rest 5 minutes at room temperature
- Repeat the vortex step 2 more times to ensure complete steroid solubility.
- The final concentration of ethanol in the sample to be added to the wells should be $\leq 5\%$. (**$\geq 1:4$ dilution with Assay Buffer is needed.**)

III. Urine:

- Urine should be diluted $\geq 1:100$ by taking one part of sample and adding 99 or more parts of Assay Buffer prior to conducting assay.
- **Normalize the sample value based on creatinine levels using our Urine Creatinine assay kit Cat NO. ZX-44110-96 in random urine specimen.**

IV. Saliva:

- Saliva samples should be frozen and thawed, then centrifuged at 14,000 rpm for 15 minutes.
- Supernatant should be diluted $\geq 1:5$ by taking one part of sample and adding 4 or more parts of Assay Buffer prior to conducting assay.

V. Tissue Culture Media:

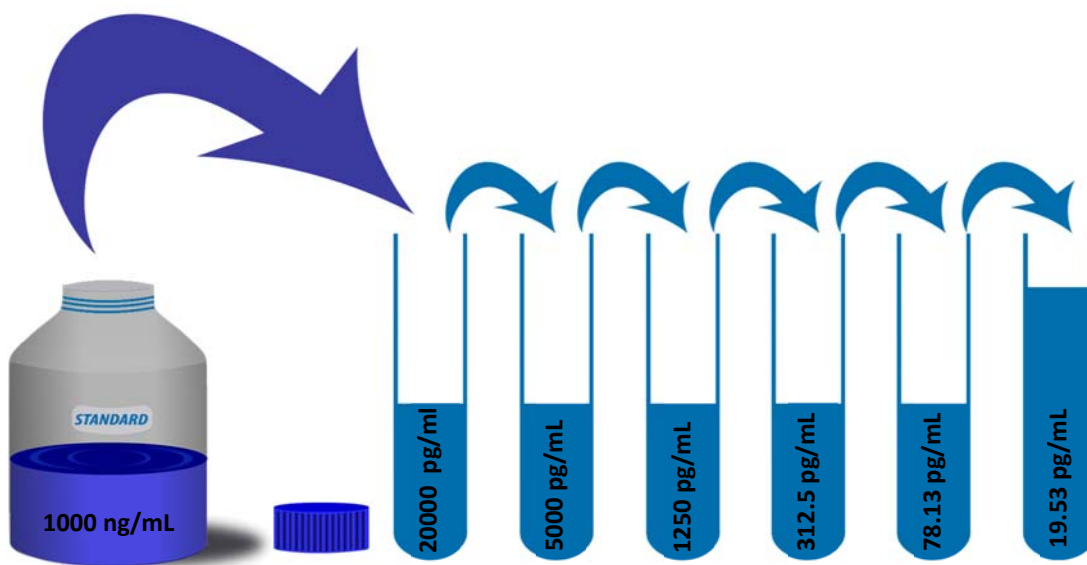
- For measuring Cortisone in tissue culture media (TCM), samples should be read off a standard curve generated in TCM. Samples may need to be diluted further in TCM. The assay has been validated using RPMI-1640.

All the samples must be used within 2 hours of preparation; otherwise, aliquots of the sample should be kept at $\leq -20^{\circ}\text{C}$ for later use.

Standard preparation

- Prepare a 1:50 dilution of Cortisone Standard with Assay Buffer (mix 10 μL of standard with 490 μL of Assay Buffer), and label as the Standard No.6 (20000 pg/mL).
- Make series of lower dilutions as described in the table.
- The Assay Buffer is used as the 0 pg/mL standard.

No.	Concentration	Material needed
Standard No.6	20000 pg/mL	10 μL Cortisone Standard + 490 μL Assay Buffer
Standard No.5	5000 pg/mL	100 μL Standard No.6 + 300 μL Assay Buffer
Standard No.4	1250 pg/mL	100 μL Standard No.5 + 300 μL Assay Buffer
Standard No.3	312.5 pg/mL	100 μL Standard No.4 + 300 μL Assay Buffer
Standard No.2	78.13 pg/mL	100 μL Standard No.3 + 300 μL Assay Buffer
Standard No.1	19.53 pg/mL	100 μL Standard No.2 + 300 μL Assay Buffer
Standard No.0	0 pg/mL	300 μL Assay Buffer



All standard must be used within 2 hours of preparation

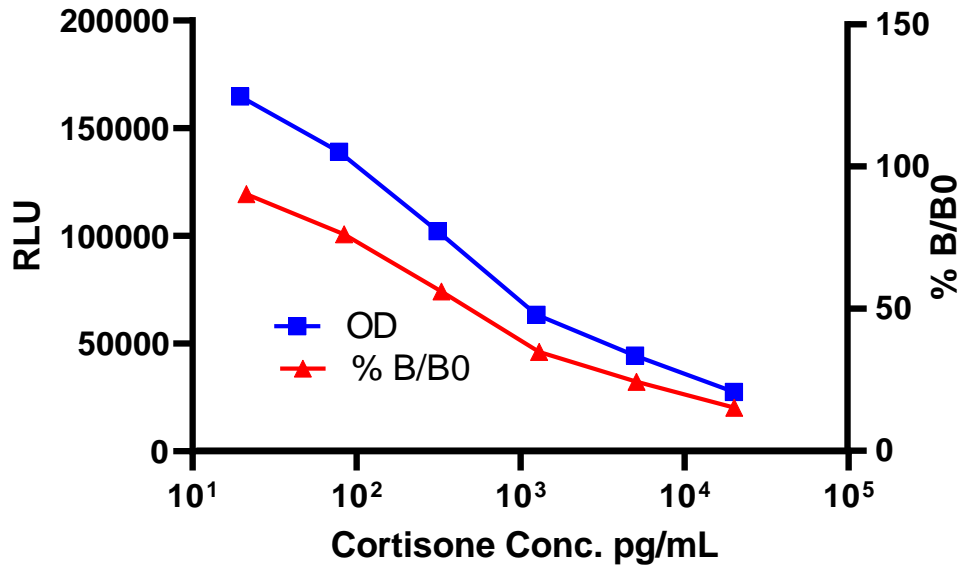
Assay Procedure

1. Pipette 100 μL of either samples or standards into duplicate wells in the plate.
 2. Pipette 100 μL of Assay Buffer into duplicate wells of the Zero standard.
 3. Pipette 125 μL of Assay Buffer into duplicate wells of the nonspecific binding (NSB).
 4. Add 25 μL of Conjugate Working Solution to each well, using a repeater pipette.
 5. Add 25 μL of Cortisone Antibody to each well except the NSB wells, using a repeater pipette.
 6. Gently tap the sides of the plate to ensure adequate mixing of the reagents.
 7. Cover the plate with the plate sealer and shake for 2 hours at room temperature. If the plate is not shaken, signals will be approximately 45 % lower.
 8. Aspirate the plate and wash each well 4 times with 300 μL Wash Buffer.
 9. Tap the plate on clean absorbent towels to dry.
 10. Add 100 μL of Chemiluminescent Substrate to each well using a multichannel/repeater pipette.
 11. Incubate at room temperature for 5 minutes without shaking.
 12. Read the luminescence generated from each well in a multimode or chemiluminescent plate reader using a 0.1 second read time per well.
- The chemiluminescent signal will decrease about 40% over 60 minutes.

Calculation

- Average the duplicate RLU readings for each standard and sample.
- Subtract the mean RLUs of the NSB from all OD values
- Create a standard curve by reducing the data using the four parameter logistic curve (4PLC) fitting routine on the plate reader
- Calculate the % B/B0 ratio.
 - **Note:** B0 is the binding for the zero standard or the maximum binding well, which represents the maximum signal from enzyme captured by the specific antibody. All other standards and samples are expressed as a percentage of this value (% B/B0).
- The concentrations obtained should be multiplied by the dilution factor to obtain sample values.

Conversion Factor: 100 pg/mL of Cortisone is equivalent to 277.6 pM



A typical standard curve of ZellX® Cortisone CLIA Assay kit
Run your own standard curves for calculation of results

Assay range

The detection limit of ZellX® Cortisone CLIA assay was determined as 16.2 pg/mL.

Sensitivity

The sensitivity of the ZellX® Cortisone CLIA assay was determined as 10.6 pg/mL.

Precision

Intra-Assay Precision (Precision within an assay): 3 human samples were tested 20 times in an assay.

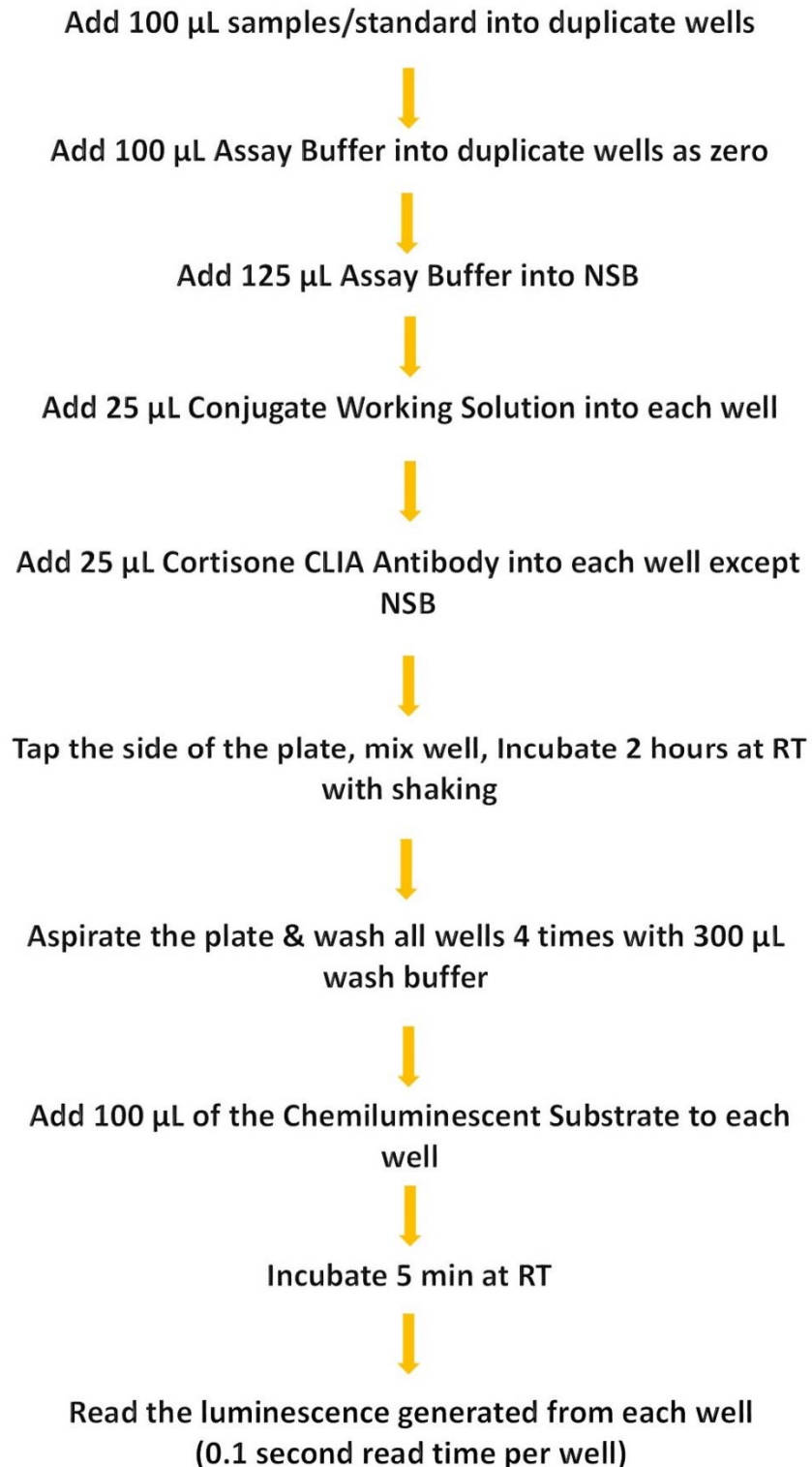
Inter-Assay Precision (Precision between assays): 3 human samples were tested in duplicate on 12 different assays over multiple days.

<i>Item</i>	<i>%CV</i>
Intra assay	13.0, 10.8, 8.6
Inter assay	9.8, 9.0, 15.1

Cross Reactivity

The following cross reactants were tested in the assay and calculated at the 50% binding point.

<i>Steroid</i>	<i>Cross Reactivity (%)</i>
Cortisone	100
5α-Dihydrocortisone	31.7
Prednisone	9.0
5β-Dihydrocortisone	4.4
11-Dehydrocorticosterone	0.62
Corticosterone	< 0.1
Cortisol	< 0.1
Estradiol	< 0.1
Progesterone	< 0.1
Dexamethasone	< 0.04

Protocol summary

References

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